

Genetic Variability within the ADA Gene and Left Ventricular Ejection Fraction

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Abstract

Background: The role of adenosine as cardio protective factor is well established. Adenosine deaminase (ADA) contributes to the control of adenosine concentration in body fluids and, as ecto-enzyme, to the regulation of adenosine receptors activity. ADA and adenosine receptors expressions have been found down regulated in heart failure. We have studied the relationship between genetic variability within the ADA gene and left ventricular ejection fraction (LVEF).

Methods: The genotypes of three polymorphic sites (SNPs) within the ADA gene have been determined in 346 patients admitted to the hospital for cardiovascular diseases. Informed consent was obtained by the patients to participate to the study that was approved by the Council of Department of Biomedicine and Prevention. The three polymorphic sites in the ADA gene are called ADA₁, ADA₂ and ADA₆. Each locus shows two alleles called ADA₁*1 and ADA₁*2, ADA₂*1 and ADA₂*2, ADA₆*1 and ADA₆*2 respectively.

Results: The joint “ADA₂*2 carrier/ADA₆*1/*1 genotype” shows a statistically significant lower value of LVEF as compared to other joint genotypes (p=0.004). Such association is statistically significant in subjects with coronary artery disease only.

Conclusions: The study of polymorphic sites of ADA gene could allow to detect subjects with higher risk of cardiac failure following infarction.

Keywords: ADA; LVEF; Genetic Polymorphism; CAD; Risk of Cardiac Failure

1. Introduction

The role of adenosine as cardio protective factor is well established [1-3]. Adenosine deaminase (ADA) and adenosine kinase activities contribute to the regulation of adenosine concentration in body fluids. Adenosine binds to specific receptors: A₁, A₂, A₃. A₁ and A₃ act through G_i protein and A_{2A} and A_{2B} through G_s protein. ADA and adenosine receptors have been found down regulated in heart failure [4].

ADA is localized on human chromosome 20q and consists of 13 exons distributed approximately on 32 Kb of DNA [5]. A number of differences among normal sequences have been found within the coding and the intronic regions of ADA gene [6]. The SNP corresponding to the presence/absence of a Taq 1 site (nt 4050-4053) in exon 1 is associated to a known functional variation and represents the basis of the common biochemical polymorphisms at ADA locus described by Spencer et al. [7]. This polymorphism is due to the presence of two common alleles ADA₁*1 and ADA₁*2: the corresponding phenotypes have enzymatic activity decreasing in the order: ADA₁1 > ADA₁ 21 > ADA₁ 2 [7]. The function of the other SNPs observed in the ADA gene has not yet been elucidated. Recent studies suggest a role of genetic variability within the ADA gene on susceptibility to coronary artery disease [8,9].

ADA is not only a cytosolic enzyme but acts as an ecto-enzyme also being present in the surface of many cell types where it regulates the extracellular concentration of adenosine and modulates the activity of receptors through interaction with other molecules [10].

We reasoned that the role of ADA in the cardiac function could be due not only to enzymatic activity i.e. the regulation of adenosine concentration, but also to ecto-enzymatic activity through interactions with adenosine receptors. Genetic variability of other sites besides the Taq 1 site in exon 1 could have an important role in this context and could influence the susceptibility to heart failure.

In the present study we have investigated three polymorphic sites (SNPs) spanning approximately 28 Kb in ADA gene in relation to the value of left ventricular ejection fraction (LVEF) an important index of cardiac function. Besides the SNP of exon 1 we have studied the Pst 1 site (ADA₂) (nt 19465-19470) of intron 2 and the MluNI site (ADA₆) (nt 31230-31235) of exon 6 [11].

2. Material and Methods

We have studied 346 patients admitted consecutively to the Hospital for Cardiovascular diseases (see Table 1). All subjects were from the White population of Rome. Venous blood samples were obtained after informed consent was acquired from all subjects to carry out the present study that was approved by the Council of Department of Biomedicine and Prevention . The data were collected a few years ago before the institution of an Ethical

Committee. All procedures followed were in accordance with the ethical standards and with the Helsinki Declaration of 1964 and its later amendments. The number of subjects is not the same in all tables owing to random missing the data for some variables. 224 blood donors were also studied as controls.

ADA genotypes were determined by RFLP-PCR. Genomic DNA was extracted from venous blood samples collected in NaEDTA using the procedure described by Kunkel [12] with slight modifications. PCR amplification was carried out as described by Hirschhorn [11]. The details of the procedure and primers have been reported in a previous paper [13]. The alleles corresponding to the presence (+) and absence (-) of the restriction sites have been indicated as allele *1 and allele *2 respectively.

Statistical analyses were carried out by commercial software (SPSS).

3. Results

Table 1 shows clinical data of the sample study.

Parameters	Proportion	Mean	S.D
Female	54.7%		
Hypertension ¹	70.7%		
Diabetes mellitus	35.2%		
Smoking habit	41.7%		
High total cholesterol ²	55.7%		
High LDL ³	55.5%		
CAD	59.2%		
Age (years)		61.8	15.5
BMI		27.4	5.2

Table 1: Clinical data in subjects with cardiovascular diseases

1 arterial tension $\geq 130/85$ mmHg

2 total cholesterol > 200 mg/dl

3 LDL >130 mg/dl

Table 2 shows the values of LVEF in relation to the polymorphic ADA loci.. No significant difference in LVEF is observed within ADA₁ genotypes. A lower value of LVEF is observed in carriers of ADA₂*2 allele as compared to ADA₂*1/*1 genotype (50.82% vs 52.79%; p=0.055). The value of LVEF is significantly lower in ADA₆*2/*2 genotype than in carriers of ADA₆*1 allele (51.40% vs 53.60%; p=0.035).

	Subjects with Cardiovascular Diseases				Controls	
	Mean LVEF	S.E.	N°	%	N°	%
ADA ₁ locus						
ADA ₁ *1/*1 genotype	52.10	0.53	302	0.88	193	0.86
Carriers of ADA ₁ *2 allele	53.10	1.34	40	0.12	31	0.14
Significance of difference	P=0.519					
ADA ₂ locus						
ADA ₂ *1/*1 genotype	52.79	0.59	205	0.60	94	0.59
Carriers of ADA ₂ *2 allele	50.82	0.88	138	0.40	65	0.41
Significance of difference	P=0.055					
ADA ₆ locus						
ADA ₆ *2/*2 genotype	51.40	0.63	233	0.67	101	0.64
Carriers of ADA ₆ *1 allele	53.60	0.80	113	0.33	57	0.36
Significance of difference	P=0.038					

Table 2: Left ventricular ejection fraction (%) in relation to the ADA polymorphic loci

Table 3 shows the value of LVEF in relation to the ADA₂-ADA₆ joint genotype. The joint genotype “ADA₂*2 carrier – ADA₆*2/*2 genotype” shows a statistically significant lower value of LVEF as compared to the other joint ADA₂-ADA₆ genotypes.

ADA ₂ -ADA ₆ joint genotype								
ADA ₂	*1/*1		*1/*1		*2 carrier		*2 carrier	
ADA ₆	*2/*2		*1 carrier		*2/*2		*1 carrier	
	(a)		(b)		(c)		(d)	
	Mean	S.D	Mean	S.D	Mean	S.D	Mean	S.D
	52.40	0.71	54.38	1.02	49.42	1.21	52.74	1.25
Total n°	151		52		80		58	
Variance analysis p=0.017								

Post hoc LSD test

c vs a p=0.021

c vs b p=0.003

c vs d p=0.038

Table 3: Left ventricular ejection fraction (%) in relation to the joint ADA₂-ADA₆ genotype.

Table 4 shows the association of the joint of “ADA₂*2 carrier-ADA₆ *2/*2 genotype” with LVEF in patients with coronary artery disease (CAD) and in patients without CAD. The association is present in patients with CAD only. In subjects with CAD multivariate statistical analyses have shown that sex, age and diabetes have no effect on the association of LVEF with the joint ADA₂-ADA₆ genotype. The effect of ADA genotype, however, is moderate as compared to that of diabetes and age (data not shown).

	“ADA ₂ *2 carriers-ADA ₆ *2/*2 joint genotype” - LVEF			Other ADA ₂ -ADA ₆ joint genotypes - LVEF			t-test for difference between means
	Mean	S.E	N*	Mean	S.E	N*	
All patients	49.42	1.21	80	52.87	0.54	261	P=0.004
Patients with CAD	45.96	1.52	52	49.69	0.72	150	P=0.015
Patients without CAD	56.04	1.44	25	57.26	0.63	107	P=0.408

Table 4: The association of LVEF with the joint “ADA₂*2 carrier-ADA₆ *2/*2 genotype”. Comparison between patients with CAD and patients without CAD.

We have also examined the relationship of the joint “ADA₂*2 carrier - ADA₆*2/*2 genotype” with LVEF divided into three classes in patients with CAD (Table 5). There is high frequency of the joint genotype “ADA₂*2 carrier - ADA₆*2/*2 genotype” in patients with LVEF ≤40 and an absence of this joint genotype in patients with LVEF > 60 (37.8% vs 0.0%, p=0.011).

		ADA ₂ *2 carrier ADA ₆ *2/*2 joint genotype	Other joint genotypes
LVEF	≤ 40	17 (37.8%)	28 (62.2%)
	40-60	35 (24.8%)	106 (75.2%)
	>60	0 (0.0%)	16 (100%)

Chi square test of independence $\chi^2 = 9.019$ df= 2 p=0.011

Table 5: The relationship of joint “ADA₂*2 carrier-ADA₆*2/*2 genotype” with LVEF divided into 3 classes in patients with CAD.

Figure 1 shows the proportion of the ADA₂*2-ADA₆*2 haplotype in relation to LVEF in subjects with CAD. The highest proportion of this haplotype is observed in subjects with LVEF ≤40 and the lowest in patients with LVEF >60.

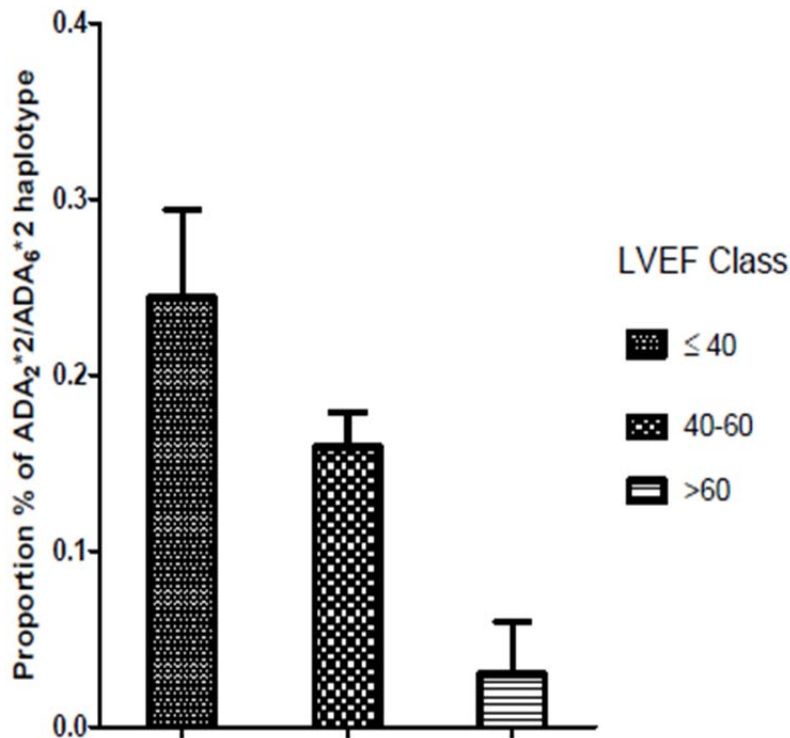


Figure 1: Proportion of ADA₂*2/ADA₆*2 haplotype in relation to LVEF class in patients with CAD

4. Discussion

The present data suggest that in the genomic area of ADA there are sites that influence LVEF. in patients with CAD. Such association is independent by other factors associated with LVEF. No correlation with the type of treatment was observed.

We have previously observed in CAD (8) a reduced proportion of ADA₂*2/ADA₆*2 haplotype as compared to controls. The positive association of this haplotype with a low LVEF suggests that the reduced proportion of this haplotype is due to a lower probability of survival and not to a protection against CAD.

Since ADA₂ is an intronic polymorphism and ADA₆ is a synonymous substitution, these DNA modifications do not change ADA protein sequence. The association of LVEF with ADA₂-ADA₆ haplotype could be due to some causal site included in this area. At present, however, cannot be excluded that genetic variability in this area influences ADA enzymatic activity and in turn adenosine concentration. It is also possible that sites in this area control aminoacid sequence of the ADA protein involved in ADA properties as ecto-enzyme.

From a practical point of view, the study of polymorphic sites of ADA gene could allow to detect subjects with higher risk of cardiac failure following infarction. Further investigation of this genomic area could be rewarding. The relatively small number of subjects examined represents the limitation of the study.

Conflict of Interest

None declared

References

1. Hori M, Kitakaze M. Adenosine, the heart, and coronary circulation. *Hypertension* 18 (1991): 565-574.
2. Kitakaze M, Hori M, Takashima S, Sato H, Inoue M, et al. Ischemic preconditioning increases adenosine release and 5'-nucleotidase activity during myocardial ischemia and reperfusion in dogs. Implications for myocardial salvage. *Circulation* 87 (1993): 208-215.
3. Khodadadi I, Vahedi MS, Abdi M, Daneshkhah N, Rahbari R, et al. Evaluation of adenosine deaminase (ADA) isoenzymes activity and tumor necrosis factor- α (TNF α) concentration in chronic heart failure. *EXCLI J* 13 (2014): 58-66.
4. Asakura M, Asanuma H, Kim J, et al. Impact of adenosine receptor signaling and metabolism on pathophysiology in patients with chronic heart failure. *Hypertens Res* 30 (2007): 781-787.
5. Wiginton DA, Kaplan DJ, States JC, et al. Complete sequence and structure of the gene for human adenosine deaminase. *Biochemistry* 25 (1986): 8234-8244.
6. Tzall S, Ellenbogen A, Eng F, Hirschhorn R. Identification and characterization of nine RFLPs at the adenosine deaminase (ADA) locus. *Am J Hum Genet* 44 (1989): 864-875.

7. Spencer N, Hopkinson D, Harris H. Adenosine deaminase in man. *Ann Hum Genet* 32 (1968): 9-14.
8. Saccucci P, Binczak-Kuleta A, Banci M et al. Coronary artery disease. A study of three polymorphic sites of adenosine deaminase gene. *Acta Cardiol* 69 (2014): 39-44.
9. Gloria Bottini F, Banci M, Saccucci P, Neri A, Magrini A, et al. A study of three polymorphic sites of ADA gene in T2D subjects with coronary artery disease. *J End Diab Res*1 (2015): 1-3.
10. Ciruela F, Saura C, Canela EI, Mallol J, Lluís C, et al. Adenosine deaminase affects ligand-induced signalling by interacting with cell surface adenosine receptors. *FEBS Lett* 380 (1996): 219-223.
11. Hirschhorn R, Yang DR, Israni A. An Asp8Asn substitution results in the adenosine deaminase (ADA) genetic polymorphism (ADA 2 allozyme): occurrence on different chromosomal backgrounds and apparent intragenic crossover. *Ann Hum Genet* 58 (1994): 1-9.
12. Kunkel LM, Smith KD, Boyer SH, et al.. Analysis of human Y-chromosome-specific reiterated DNA in chromosome variants. *Proc Natl Acad Sci* 74 (1977): 1245-1249.
13. Sebastiani GD, Bottini N, Greco E, et al.. A study of Adenosine-Deaminase genetic polymorphism in rheumatoid arthritis. *Int J Immunopathol Pharmacol* 23 (2010): 791-795.



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