

## Prevalence of Intestinal Parasites in Vegetables Sold in Some Local Markets in Port-Harcourt, Rivers-State, Nigeria

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### Abstract

A total of 492 vegetable leaves samples made up of cabbage (27), bitter leaves (50), garden egg leaves (44), green leaves (66), green onion (47), lettuce (51), pumpkin leaves (111), parsley (26) and water leaves (70), bought randomly from various local markets in Port-Harcourt was analyzed for the presence of some parasite stages. About 145(29.5%) out 492 samples examined were contaminated with various parasite stages. Green leaves had the highest level of contamination (40.9%), followed by bitter leaves (38.8%), while cabbage recorded the lowest (7.4%). Out of 202 parasite stages identified, the eggs of *Ascaris lumbricoides* were the most predominant 39 (19.3%), followed by the cysts of *Entamoeba histolytica* 31 (15.3%), whereas the oocysts *Cryptosporidium parvum* and eggs of *Strongyloides stercoralis* had the least 4 (1.9%). Green leaves and pumpkin leaves harboured 8 (80.0%) and 7 (70%) respectively out of 10 different types of parasites obtained in this study; cabbage and parsley had only 3 (30.0%). Pumpkin leaves recorded the highest level eggs of *Ascaris lumbricoides* 18(30%), eggs of *Fasciola* species 10 (16.7%), cysts of *Entamoeba histolytica* 8 (50.0%), eggs of *Giardia lamblia* 7 (11.7%), whereas cabbage overall had the lowest percentage of parasites with *Ancylostoma duodenale* 2 (50.0%); eggs of *Toxoplasma gondii* 1 (25.0%) and eggs of *Strongyloides stercoralis* 1 (25.0%). The present study revealed the potential hazard of vegetable leaves sold in some local markets in Port – Harcourt. Though the vegetables are properly de-contaminated after sales at the point of preparation/cooking, it is also very necessary for the de-contamination to be done before sales. There is

need for monitoring by educating both vendors and consumers on the good sanitary practices during processing displaying and sale of the vegetable products and possible danger of contaminated and spoiled vegetables.

**Keywords:** Prevalence; Parasites; Vegetables; Port – Harcourt

## 1. Introduction

Fresh vegetables are regarded as important part of a healthy diet. They are horticultural products, having tissues with continuing metabolism and subject to respiration, water loss and cell softening throughout the post – harvest period [1]. Vegetables are a good source of antioxidants and phytonutrients. They are low in calories and rich in complex carbohydrates, vitamins and minerals. In many countries, such leafy plants are eaten raw or lightly cooked to preserve taste and this practice may also favour the likelihood of food borne parasitic infections [2]. Pathogens on edible plants present a significant potential source of human illness. The pathogenic microorganisms which reside on the soil and in the intestinal tract of humans and animals are more likely to contaminate vegetables through faeces, sewages, untreated irrigation water or surface water [3]. Contamination can also take place during handling of the product, processing and transportation [4, 5]. Bacteria, viruses and parasites on vegetables have been linked with diseases [6, 7] and the most common intestinal parasites on raw vegetables are protozoans, nematodes, cestodes and trematodes [8, 9]. Intestinal parasitic infections are among the major public health and socio-economic concerns that affect well –being of the poor in developing countries. It has been estimated that *Ascaris lumbricoides*, hookworm and *Trichuris trichuria* infect 1.050 million, 1.300 million and 1.050 million people worldwide respectively, while schistosomiasis affects over 200 million people [10]. *Entamoeba histolytica* and *Giardia lamblia* are also estimated to infect about 60 million and 200 million people worldwide respectively [11].

## 2. Materials and Methods

A total of 492 vegetable leaves samples made up of cabbage leaves (27), bitter leaves (50), garden egg leaves (44), green leaves (66), green onion (47), lettuce (51), pumpkin leaves (111), parsley (26) and water leaves (70) were bought randomly from various local markets in Port-Harcourt Rivers-State from February –December 2015. About 250 g of each vegetable samples purchased was sliced into small pieces, packaged into a plastic bag and washed with 1000 ml of physiological saline solution (0.95% NaCl). The washing water/saline was left overnight. After the removal of leaves, about 400 ml of supernatant was transferred into 5ml test tubes and centrifuged at 2000 g for 20 minutes. After centrifugation, the supernatant was discarded and the sediment examined under light microscope, using x10 and x40 objectives for parasite stages (cysts, oocysts, eggs or larvae) after adding a drop of lugol iodine and the parasite stages identified. Positive samples were recorded and developmental stages recorded.

## 3. Results

The frequency of distribution of different infected vegetable leaves consumed in Port-Harcourt is shown in Table 1. Out of 492 vegetables leaves examined, 45 (29.5%) harboured parasite stages. Green leaves (*Celosta argentea*) had the highest level of intestinal parasite contamination 27 (40.9%), followed by bitter leaves (*Vernomia amygdlima*) 19 (38.0%), while cabbage (*Brassica oleracae*) samples recorded the lowest level of contamination 2 (7.4%). The

eggs of *Ascaris lumbricoides* were the most predominant 39 (19.3%), followed by cysts of *Entamoeba histolytica* 31(15.3%), whereas the oocysts of *Cryptosporidium parvum* and eggs of *Strongyloides stercoralis* recorded the lowest percentage of occurrence of 4 (1.9%) as shown in Table 2. Table 3, shows a seasonal variation of the density of parasitic infections in the vegetables examined. The highest rate was recorded in the months of July 31 (15.3%), August 24(11.9%), followed by the month of June 20 (9.9%); while November got only 14 (6.9%). Table 4 shows the distribution of intestinal parasites in vegetable leaves consumed in Port-Harcourt. Out of 10 different parasites identified during this survey, green leaves harboured 8 (80%) different parasites, followed by bitter leaves and pumpkin leaves (*Telfairi occidentalis*) 7 (70%) each, where as cabbage and parsley (*Petroselinum crispum*) recorded the least 3 (30%). The majority of parasites found in this study was identified from pumpkin leaves samples with *Ascaris lumbricoides* 18 (30%), eggs of *Fasciola* species 10 (16.7%), eggs of *Taenia* species, cysts of *Entamoeba histolytica* 8 (50.0%), eggs of *Giardia lamblia* 7 (11.7) where as cabbage recorded the least number of parasites. Eggs of *Toxoplasma gondii* 1 (25%), eggs of *Strongyloides stercoralis* 1 (25.0%) and eggs of *Ancylostoma duodenale* 2 (50%).

Vegetable	Leaves	Number examined	Number positive (%)
Cabbage	( <i>Brassica oleraceae</i> )	27	2 (7.4)
Bitter leaf	( <i>Vernonia amygdalina</i> )	50	19 (38.0)
Garden egg leaves	( <i>Solanum manocarpum</i> )	44	15 (34.1)
Green leaves	( <i>Celosta argentea</i> )	66	27 (40.9)
Green onion	( <i>Allium wakegi</i> )	47	4 (8.5)
Lettuce	( <i>Letuca sativa</i> )	51	12 (23.5)
Pumpkin leaves	<i>Telfairi occidentalis</i> )	111	40 (36.0)
Parsley	( <i>Petroselinum crispum</i> )	26	4 (15.4)
Water leaves	( <i>Talilum triangulare</i> )	70	22 (31.4)
Total		492	145 (29.5)

**Table 1:** Frequency of distribution of different infected vegetable leaves consumed in Port -Harcourt City, Rivers-State.

Parasites	Number of parasites	Occurrence
<i>Ancylostoma duodenale</i>	25	12.3
<i>Ascaris lumbricoides</i>	39	19.3
<i>Cryptosporidium parvum</i>	4	1.9
<i>Entamoeba coli</i>	27	13.4
<i>Entamoeba histolytica</i>	31	15.3

<i>Gardia lambia</i>	24	11.9
<i>Fasciola species</i>	19	9.4
<i>Taenia species</i>	22	10.9
<i>Toxoplasma gondii</i>	7	3.5
<i>Strongyloides stercolaris</i>	4	1.9

**Table 2:** Percentage (%) of occurrence of Intestinal Parasites in Vegetable Leaves Consumed in Port-Harcourt, Rivers – State.

Examination Period	Number of Parasites (%)
February 2015	13 (6.4)
March 2015	15 (7.4)
April 2015	16 (7.9)
May 2015	18 (8.9)
June 2015	20 (9.9)
July 2015	31 (15.3)
August 2015	24 (11.9)
September 2015	15 (7.4)
October 2015	19 (9.4)
November 2015	14 (6.9)
December 2015	17 (8.4)
February 2015 – December 2015	202 (100)

**Table 3:** Seasonal Variation of Parasites Identified in the Vegetable Leaves Consumed in Port - Harcourt, Rivers - State; From February 2015 to December 2015.

Vegetable leaves	<i>Ancylostoma duodenale</i>	<i>Ascaris lumbricoïdes</i>	<i>Cryptosporidium Parvum</i>	<i>Entamoeba coli</i>	<i>Entamoeba histolytica</i>	<i>Giardia lambia</i>	<i>Fasciola species</i>	<i>Taenia species</i>	<i>Toxoplasma gondii</i>	<i>Strongyloides stercolaris</i>
Cabbage ( <i>B oleracae</i> )	2 (50.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (25.0)	1 (25.0)
Bitter leaves ( <i>V amygdalina</i> )	2 (10.0)	3 (15.0)	2 (10.0)	4 (20.0)	6 (30.0)	0 (0.0)	1 (5.0)	2 (10.0)	0 (0.0)	0 (0.0)
Garden egg leaves ( <i>S.monocarpum</i> )	0 (0.0)	5 (21.7)	2 (8.7)	9 (39.1)	2 (8.7)	3 (13.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (8.7)
Green leaves ( <i>C. argentea</i> )	4 (12.1)	5 (15.1)	0 (0.0)	6 (18.1)	6 (18.1)	5 (15.1)	2 (6.1)	4 (12.1)	1 (3.0)	0 (0.0)

Green onion ( <i>A. wakegi</i> )	1 (12.5)	2 (25.0)	0 (0.0)	4 (50.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	0 (0.0)	1 (12.5)
Lettuce ( <i>L. sativa</i> )	3 (18.8)	0 (0.0)	0 (0.0)	0 (0.0)	4 (25.0)	3 (18.8)	6 (37.5)	0 (0.0)	0 (0.0)	0 (0.0)
Pumpkin leaves ( <i>T. occidentalis</i> )	4 (6.7)	18 (30.0)	0 (0.0)	0 (0.0)	8 (50.0)	7 (11.7)	10 (16.7)	10 (16.7)	3 (5.0)	0 (0.0)
Parsley ( <i>P. crispum</i> )	1 (16.7)	0 (0.0)	0 (0.0)	0 (0.0)	3 (50.0)	0 (0.0)	0 (0.0)	0 (0.0)	2 (33.3)	0 (0.0)
Water leaves ( <i>T. triangulare</i> )	8 (25.0)	6 (18.8)	0 (0.0)	4 (12.5)	2 (6.3)	6 (18.8)	0 (0.0)	6 (18.8)	0 (0.0)	0 (0.0)

**Table 4:** Percentage distribution of intestinal parasites in vegetable leaves consumed in Port – Harcourt.

#### 4. Discussion

The risk of infection with intestinal parasites to the population is increasing because the contaminated vegetables are sometimes eaten undercooked to retain the natural taste and preserve heat labile nutrient [12]. During this study, about 145 (29.5%) out of 492 Vegetable samples examined in Port-Harcourt city were contaminated with different parasite stages. The evidence of this moderate parasite score densities among vegetable samples examined indicates high transmission risk of such parasites to the inhabitants of the city. Analysis during this survey showed that all types of vegetables analyzed were contaminated with various infestation rates as: Green leaves recorded (40.0%), bitter leaves (38.0%), pumpkin leaves (36.0%), Garden egg leaves (34.0%), water leaves (31.4%), lettuce (23.5%), parsley (15.4%), green onion (8.5%) and cabbage (7.4%). This is in contrast to the reports of Wafa and Megrin et al. [13]. The contamination of all vegetables can be attributed to the handling technique [14], also to dirty and untrimmed nails and poor hand - washing habits [14]. Stages of parasites belonging to 10 genera known as: ova of *Ascaris lumbricoides*, cysts of *Entamoeba histolytica* and *Entamoeba coli*, trophozoites of *Giardia lamblia*, eggs of *Taenia* species, eggs of *Fasciola* species, cysts of *Toxoplasma gondii* and eggs of *Strongyloides stercoralis* identified in this study had previously been recovered from leafy vegetables in Riyadh [13]. *Ascaris lumbricoides* (19.3%) was the most prevalent parasites; which is supported by the fact that *Ascaris lumbricoides* is more common in tropical regions [16], and also in accordance with the reports that found *Ascaris lumbricoides* being the most common parasites in vegetables [17]. These findings are in accordance with the reports of Uneke [9]; Damen et al. [18] and Alade et al. [19]. But in contrast to the reports that found *Entamoeba coli* and *Ancylostoma duodenale* as the most frequently encountered parasites on vegetables [20]; their presence may be due to the use of excreta as manure commonly practiced by vegetables farmers. Also *Ascaris* eggs are more resistant than other intestinal parasites to most types of inactivation processes. A temperature of 20-30°C, pH plays a significant role in the survival of *Ascaris* eggs; however above 50°C temperature becomes the most important factor for their survival [21]. *Entamoeba histolytica* (15.3%) and *Entamoeba coli* (13.4%) were second and third predominant parasites identified in this study, which is in accordance with reports of the study carried out by Al- Binali et al. but in contrast with the reports of Vuong et al. [23]; Darchenkova et al. [24]. Their noticeable presence could be attributed to the contamination of fresh vegetables during production, harvest, packing and distribution [25], contamination through soil, faeces (human and animal origin), water irrigation and application of manures or sewage [26, 27]. Besides, cysts of intestinal parasites have been found to adhere more easily to the surface of vegetables. *Entamoeba coli* is a non - pathogenic parasite (denotes a faecal contamination), but a recently recognized *Entamoeba coli* strain was reported to produce high toxins that cause kidney damage as septicaemia or blood poisoning. Symptoms can include diarrhea,

chills, headaches and high fever and in some infections can lead to death even with medical intervention [28]. The increased demand, global sourcing and rapid transport of food, especially soft fruits and vegetables enhance both likelihood of surface contamination and survival of transmissible stages of parasites pathogenic to man. *Cryptosporidium parvum* is well known to be a causative agent of prolonged diarrhea among HIV/AIDS patients, but its identification could be attributed to the fact that as a zoonotic parasite can be transmitted by water borne and food borne routes. That means contact between fresh vegetables and some infected animals (public, pet or wild) could had been another source of these intestinal parasites [29].

Moderate presence of *Giardia lamblia* and *Strongyloides stercoralis* is similar to the reports of Ali et al. [20]. Evidence has also indicated that infection with *Giardia lamblia* could cause malabsorption, chronic diarrhea and long term growth retardation in children [30]; While *Strongyloides stercoralis* causes Strongyloidiasis which presents no destructive picture and contamination of vegetables becomes obvious when the eggs of *Strongyloides stercoralis* remain quiescent in the film of contaminated soil and there is contact with the vegetation (vegetables). The presence of *Toxoplasma gondii*, *Taenia* species and *Fasciola* species in this study was not in agreement with the reports of Nyarango et al. [31]. The water-food connection for parasites zoonoses is complex, with faeces as a major vehicle for many environmental transmission stages. The transmissible stages can contaminate food or vegetables directly voided in faeces, or indirectly water is a major conduct for these parasites by the use of contaminated water which in turn can contaminate vegetable. Contamination can also occur when vegetables are rinsed in parasites contaminated water. *Toxoplasma gondii* has been associated with waterborne diseases. *Taenia* and *Fasciola* species identification is a possible justification of the use of animal manure as fertilizers and waste water effluent for irrigating aquatic and semi-aquatic vegetable crops [32].

This study was conducted during a period of 10 months (February 2015-December 2015). It was observed that parasite density rates were higher in the month of July, August, June and October being the rainy season in Port - Harcourt Rivers-State. The reason of this finding is characterized by the fact that rain splashes, irrigation or river flush of contaminated soil or poor sewage disposal during heavy rainfall will deposit contaminated soil on the surface of leaves of vegetables. These findings are in contrast to the reports by Ali et al and Monger et al. [20], but similar to the reports that recorded higher parasite rates in rainy season [33]. February recorded only 6.4% parasite rate and the reason was not well known.

Despite the widespread incidence of these parasites, the non-acute nature of these infections has contributed to the perception of intestinal parasites being common, but usually unimportant to the public health community. Over the years, several estimates of global and regional prevalence and number of infections by intestinal parasites have been made, and these estimates project similar pictures [34, 35]. *Ascaris* still remain the most common intestinal parasite and the most prevalent in Sub-Saharan Africa and India [36]. Results of this present study showed significant rate of parasite in vegetables in Port-Harcourt which is of serious public health concern. Results of this work indicate poor hygienic conditions of the vegetables and the consumers are at risk of contracting food borne diseases. Though the vegetables are properly de-contaminated after sales at the point of preparation/cooking, it is also very necessary for

the de-contamination to be done before sales. Therefore measures should be taken to spread awareness among vendors about the safe and hygienic practices and government agencies should take initiative to lay out guidelines for selling of fresh fruits and vegetables.

## 5. References

1. Obetta SE, Nwakonobi TU, et al. Microbial Effects on Selected Stored Fruits and Vegetables Under Ambient Conditions in Makurdi, Benue State, Nigeria. *Research Journal of Applied Sciences, Engineering and Technology* 3(2011): 393-398.
2. Ozlem, Sener. Survey of Fresh Vegetables for Nematodes. *Journal of Association of Analytical Chemistry* 6(2005): 613-615.
3. Feng P, Weagant SD, et al. Enumeration of *Escherichia coli* and the Coliform Bacteria. *Bacteriological Analytical Manual* (8<sup>th</sup> Edn) revision A.U.S. Food and Drug administration, college park, M.D (2002).
4. Johannessen GS, Loncarevic S, et al. Bacteriological Analysis of Fresh Produce in Norway. *International Journal of Food Microbiology* 77 (2002): 199-204.
5. Maizes RM, Yazdanbaksh M. Immune Regulation by Helminth Parasites; Cellular and Molecular Mechanism''. *Nat. Rev. Immunology* 3 (2003): 733-744.
6. Rajvanshi A. Bacteria load on Street Vended Salad in Jaipur City, India. *Internet Journal of Food Safety* 12 (2010): 136-139.
7. De Roever C. Microbiological Safety Evaluations and Recommendations on Fresh Production. *Food Control* 19 (1999): 727-733.
8. Doyle ME. Food Borne Parasites: A Review of the Scientific Literature. Food Research Institute. University of Wisconsin, Madison (2003): 1-28.
9. Uneke CJ. Potential for Geohelminth Parasites Transmission by Ray Fruits and Vegetables in Nigeria Implication for a Risk Profile. *Journal of Nutritional and Environmental Medicine* 16 (2004): 59-68.
10. World Health Organization. Expert Committee Prevention and Control of Schistosomiasis and Soil Transmitted Helminthiases. WHO Technical Report Series 912 (2002): 1-57.
11. Murray PR, Rosenthal KS, et al. *Medical Microbiology*, 4<sup>th</sup> Edn. London, Mosby (2002): 618-761.
12. Slifco TS, Smith HV, et al. Emerging Parasite Zoonoses Associated with Water and Food. *International Journal for Parasitology* 30 (2000): 1379-1393.
13. Wafa AL, Al-Megrin. Prevalence of Intestinal Parasites in Leafy Vegetables in Riyadh, Saudi Arabia. *International Journal of Tropical Medicine* 5 (2010): 20-23.
14. Mustafa U, Adnan S, et al. Environmental Pollution with Soil Transmitted Helminthes in Sanliurfa, Turkey. *Mem. Inst. Oswaldo Cruz Rio de Janeiro* 96 (2001): 903-909.
15. Dongue AR, Deshmukh PR, et al. An Approach to Hygiene Education Among Rural Ludiam School Children. *Online Journal of Health and Allied Sciences* (2008).

16. Stepek G, Buttle DJ, et al. Human Gastrointestinal Nematode Infections: Are New Control Methods Required? *Int J Exp Patho* 87 (2006): 324-341.
17. Srikanth R, Naik D. Health Effects of Water Reuse for Agriculture in the Suburbs of Asmara City, Eritrea. *Inter J Occup Environ Health* 10 (2004): 284-288.
18. Damen JG, Benwat EB, et al. Parasitic Contamination of Vegetables in Jos, Nigeria. *Annals of African Medicine B*: (2007): 115-118.
19. Alade GO, Alade TO, et al. Prevalence of International Parasites in Vegetables Sold in Ilorin, Nigeria. *American, Eurasian J Agric and Environ Sci* 13 (2013): 1275-1282.
20. Ali M, Al-Bimali S, et al. The Prevalence of Parasites in Commonly Used Leafy Vegetables in South Western Saudi Arabia. *Saudi Arabia Med J* 27 (2006): 613-616.
21. Pescon BM, Barrios JA, et al. The Effects of Temperature, pH and Ammonia Concentration on the Inactivation of Ascaris eggs in Sewage Sludge. *Water Res* 41 (2007): 2893-2902.
22. Bethony J, Albonico M, et al. Soil Transmitted Helminth Infections: Ascariasis, Trichuriasis and Hookworm. *The Lancet* 67 (2006): 1521-1532.
23. Vounou BG, Chitwood MB, et al. *Introduction to Nematology* 3 (2007): 85-90.
24. Darshenkova Payne WL, Jackson GJ. *Journal of Occupation Environmental Health* 1 (2006): 284-288.
25. Bartz JA, Wei CI. *The Influence of Bacteria Post-Harvest Physiology and Pathology of Vegetables*. 2ed., Marcel Dekker, Inc, New York (2003): 519-541.
26. Olayemi AB. Microbiological Hazards Associated with Agricultural Utilization of Urban Polluted Water. *International. J Environ Health Res* 7 (2007): 149-154.
27. Ward EF, Irvring LG. Virus Survival on Vegetables Spray Irrigated with waste water. *Wat Res* 21(1987): 57-63.
28. Tsado EK, Adesina OA, et al. A Survey on Bacterial Load of Selected Fruits and Leafy Vegetables in Minna Metropolis of Niger State, Nigeria. *Journal of Animal Production Advances* 3 (2013): 6-11.
29. Evans MR, Gardner D. Cryptosporidiosis Outbreak with Educational Farm Holiday. *Commun Dis Rep CDR Rev* 6 (1996): R50-R51.
30. Fraser DN, Bilenko N, et al. Giardia lamblia Carriage in Isreali; Bedouin Infants : Risk Factors and Consequences *Clinical Infectious Diseases* 30 (2000): 419-424.
31. Nyarango RM, Aloo PA, et al. The Risk of Pathogenic Intestinal Parasite Infections in Kisii Municipality, Kenya. *BMC Public Health* 8 (2008): 237.
32. World Health Organization. Control of Food-Borne Trematode Infections. WHO Technical Report 849 (1995): 1-157.
33. Robertson LJ, Gjerde B. Occurrence of Parasites on Fruits and Vegetables in Norway. *J. Food Prot* 61 (2001):1793-1798.
34. Erdogrul O, Sener H. The Contamination of Various Fruits and Vegetables with Enterobius vermicularis, Ascais eggs, Entamoeba histolitica cysts and Giardia cysts. *Food control* 16 (2005): 559-562.



35. Hotez PJ, Bottazzi ME, et al. The Neglected Tropical Disease of Latin America and the Caribbean: A Review of Disease Burden and Distribution and a Roadmap for Control and Elimination. *Plos Negi Trop Dis* 2 (2008): e300.
36. Bethony J, Brooker S, et al. Soil Transmitted Helminth Infections: Ascariasis, Trichuriasis and Hookworm. *Lancet* 367(2006): 1521-1532.



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